



Research Article

EVALUATION OF THE HEPATOPROTECTIVE AND ANTIOXIDANT EFFECT OF  
*BERBERIS ASIATICA* AGAINST EXPERIMENTALLY INDUCED LIVER INJURY IN RATS

BRIJESH.K.TIWARI<sup>1</sup>, R.L KHOSA<sup>2</sup>

<sup>1</sup>Translam Institute of Pharmaceutical Education and Research, Meerut,-25001-India, <sup>2</sup>Department of Pharmacy, Bharat Institute of Technology Meerut,-250002 India. Email: transbrijesh@gmail.com

ABSTRACT

*Berberis asiatica* roots have been used for the treatment of affection of eyes, skin, rheumatism and jaundice could be a substitute for endangered species *Berberis aristata* exhaustively used by industries and in ayurvedic system of medicine. The present investigation describes hepatoprotective and antioxidant activities of dried aerial parts of *Berberis asiatica*, aqueous (AqBA) and methanol (MeBA) extracts, against CCl<sub>4</sub> -induced hepatic injury. Hepatic injury was achieved by injecting 2ml/kg s.c of CCl<sub>4</sub> in equal proportion with olive oil. AqBA and MeBA at dose levels of 200 and 300 mg/kg offered significant ( $P<0.001$ ) hepatoprotective action by reducing the serum marker enzymes like serum glutamate oxaloacetate (SGOT), serum glutamate transaminase (SGPT). They also reduced the elevated level of serum alkaline phosphatase (ALP) serum acid phosphatase (ACP) and serum bilirubin. Reduced enzymic and non enzymic antioxidant levels and elevated lipid peroxide level were restored to normal by administration of MeBA and AqBA. Histopathological studies further confirmed the hepatoprotective activity of these extracts when compared with CCl<sub>4</sub> treated control groups. The result obtained were compared with silymarin (100mg/kg; p.o), the standard drug. In conclusion MeBA extract at (200mg/kg, p.o) showed significant  $p<0.001$  hepatoprotective activity similar to that standard drug, silymarin.

**Keywords:** *Berberis asiatica*; antioxidant activity; antihepatotoxic activity, free radicals

INTRODUCTION

Free radicals are reactive oxygen species (ROS) are inevitably generated due to incomplete reduction of O<sub>2</sub> in electron transfer reaction as a byproduct of biological reactions. The reactive oxygen species (ROS) such as superoxide anion radical (O<sub>2</sub><sup>-</sup>), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and hydroxyl radical (·OH) have been implicated in the pathophysiology of various clinical disorders, including ischemia, reperfusion injury, atherosclerosis, acute hypertension, haemorrhagic shock, diabetes mellitus and cancer.<sup>1</sup> They play an important role in the inflammation process after intoxication with by ethanol, carbon tetrachloride or carrageenan.<sup>2,3,4</sup> It is well known that, free radicals are the reactive species derived from them cause damage through mechanisms of covalent binding and lipid peroxidation with subsequent tissue injury.<sup>5</sup> Scavenging of free radicals by antioxidants could reduce the fibrosis in the tissue. <sup>6</sup> Thus Antioxidant agents of natural origin have attracted special interest because they can protect human body from free radicals.<sup>7</sup> numerous medicinal plants and their formulations are used for liver disorders in ethno medical practices as well as in traditional systems of medicine in India.<sup>8</sup>

*Berberis asiatica* Roxb. (Berberidaceae) is a very common substitute to "*Daruharidra*," that is *Berberis*

*aristata* DC which is used in ayurvedic system of medicine. Being an important medicinal plant, it is used extensively for treating variety of ailments. That is affection of eyes, skin diseases, jaundice and rheumatism.<sup>9-11</sup>

The major alkaloid of this plant is reported to be berberin.<sup>12-13</sup> Ethnomedicinal investigations revealed that the tribal kumaun region use the decoction of the root for treating the eye troubles and boils.<sup>14</sup> The extensive use of *Berberis aristata* by different pharmaceutical industries coupled with recent revival of interest in herbal medicines have led to ever increasing demand of this species. It as therefore becomes essential to search for a possible substitute for this species. The present investigations of methanol extract of *Berberis asiatica* aerial parts studied for its possible anti hepatotoxic and antioxidant action in rats and aimed to evaluate the relationship between liver protective effects and antioxidant activity.

MATERIAL AND METHODS

Plant material

The aerial parts of *Berberis asiatica* Roxb. (Berberidaceae) were collected from Pithoragarh district of Uttarakhand during January 2007. The plants were identified by Dr. H.B Singh of National

Institute of Science communication and Information recourses (NISCAIR), New Delhi, India.

### Preparation of extract

The dried ground aerial parts of *Berberis asiatica* were extracted by methanol and water separately using Soxhlet apparatus. The extracts obtained were dried under reduced pressure using rotary evaporator to get the crude aqueous extract (21%) and methanol extract (24 %) respectively.

### Phytochemical screening

The known quantity of dried powdered drug was extracted in soxhlet with pet ether, methanol and water successively (Table.1) was tested for different constituents (Peach and Tracy, 1955) viz. steroids and triterpenoids (Liebermann-Burchard reaction), flavonoids (Shinoda test), alkaloids (Mayer's reagent), tannins (Ferric chloride test) and sugar (Fehling solution test). The phytochemical investigations revealed that triterpenoids and alkaloids were present in methanol and water soluble parts, resin and sugar in pet ether fractions and tannins were present in water soluble parts.

### Animals

Wistar albino rats (150-200g) of either sex roughly of the same age (8-10 weeks) were used for the present studies. They were housed in clean polypropylene cages (3 in each) and maintained in standard laboratory condition at ambient temperature (25±2°C) with relative humidity (55-64%) and light and dark conditions (12/12h). They were provided with rat chow diet (Hindustan Liver limited Kolkata) and free access of drinking water *ad libitum*. The experiments and procedure used in the study were approved by Institutional animal ethical committee (IAEC).

### Chemicals

Silymarin, (Sigma chemicals, USA), C<sub>2</sub>H<sub>5</sub>OH, disodium hydrogen phosphate (Na<sub>2</sub>HPO<sub>4</sub>), Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), dihydrogen potassium phosphate anhydrous (KH<sub>2</sub>PO<sub>4</sub>) and thiobarbituric acid were purchased from Merck India Ltd, Mumbai India.

### Assessment of hepatoprotective activity

This was described recently (Tiwari et al., 2008). Briefly the animals were divided into six groups of six animals each. Group I served as normal control were administered p.o., a single daily dose of 0.5% Tween 80 (1ml) on all 5 days and olive oil (1ml/kg) on days 2 and 3. Group II served as induction control, animals were administered a single daily dose of 0.5% Tween 80 (1ml) p.o., on all 5 days and on 2 and 3 day they were administered CCl<sub>4</sub>, 2mL/Kg s.c., (1:1) dilution with olive oil (Lin et al., 1998) on 5 day. Group III-VI, served as *Berberis asiatica* treated groups were administered AqBA and MeBA extracts (200 and 300mg/kg, p.o.) all 5 days and a single dose of CCl<sub>4</sub>

(2mL/Kg) s.c., on days 2 and 3, 30 min after extracts administrations and group VII Silymarin, the known hepatoprotective compound (100mg/kg.p.o.) for 5 days and a single dose of CCl<sub>4</sub> (2mL/Kg) s.c., on days 2 and 3, 30 min after silymarin administration. The food was withdrawn on preceding night of the experiment. On 5<sup>th</sup> day all the animals were sacrificed by mild ether anesthesia. Blood samples were collected from heart of each animal. Serum was separated for the estimation of the biochemical markers and liver was dissected out for the determination of antioxidant activity for histology studies liver tissue was collected.

**Table 1: Phytochemical screening of extracts.**

Phytoconstituents	Qualitative abundance		
	Pet. ether	Methanol	Water
Antraquinones	-	-	-
Soluble tannins	+	++	++
Condensed tannins	+	++	+
Flavonoids	-	++	-
Alkaloids	+	+++	++
Indole alkaloids	-	++	+
Steroidal alkaloids	-	++	+
Saponins	+	+	-
Glycosides	-	-	-
Resins	+	-	-
Terpenoids	-	++	+

Levels of phytoconstituents were qualitatively determined based on chemical groups and thin layer chromatography on the following scale, - absent +present at low level, ++ present at moderate level, +++ present at high level.

### Determination of enzyme level

The activities of serum glutamate pyruvate transaminase (SGPT), and serum glutamate oxaloacetate transaminase (SGOT) estimation of serum ALP, serum Bilirubin [Total and direct] were assayed by the reported methods<sup>18, 19, 20</sup> and carried out to assess the acute hepatic damage caused by CCl<sub>4</sub>.

### Assessment of antioxidant activity

Liver was excised washed thoroughly in 0.86% ice cold saline to remove the red blood cell. They were then gently blotted between the folds of filter paper and weighed in an analytical balance. Then it was suspended in 10 % (w/v) in ice cold buffer 0.05M then phosphate buffer cut into small pieces and homogenized using a polytron homogenizer at 20°C. The homogenate was centrifuged at 3000 rpm for 20 min to remove the cell debris, unbroken cells, nuclei erythrocytes and mitochondria. The supernatant was used for the estimation of enzymic and non enzymic antioxidants like superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), activities and the level of lipid peroxidation as described by the methods<sup>21-24</sup>

### Histopathological Examination

The liver tissue was collected and immediately fixed in 10% formalin solution, dehydrated in gradual ethanol

(50-100%), cleared in xylene and embedded in paraffin. sections (4-5 $\mu$ m) were prepared and then stained with hematoxylin and eosin dye for photo microscopic observations.

### Statistical analysis

The data represents mean  $\pm$  SD. Results were analyzed statistically by one way ANOVA followed by Bonferroni's multiple comparison test between the data of control and treated groups using SPSS software (students' version ), the minimum level of significance was set up at  $p < 0.05$  or less.

### RESULTS

On preliminary phytochemical analysis various fraction of *Berberis asiatica* have shown the presence of steroids and triterpenoids, flavonoids, alkaloids tannins and sugar as represented in table.1

As shown in table 2 activities of serum GPT, GOT, acid and alkaline phosphatase were markedly elevated in CCl<sub>4</sub> treated animals compared to normal, indicating liver damage. Administration of methanol extract at 200mg/kg remarkably prevented CCl<sub>4</sub> elevation of serum enzymes. The AqBA have shown nearly same effect at higher dose (300mg/kg).(fig 1(a) and fig(1b)

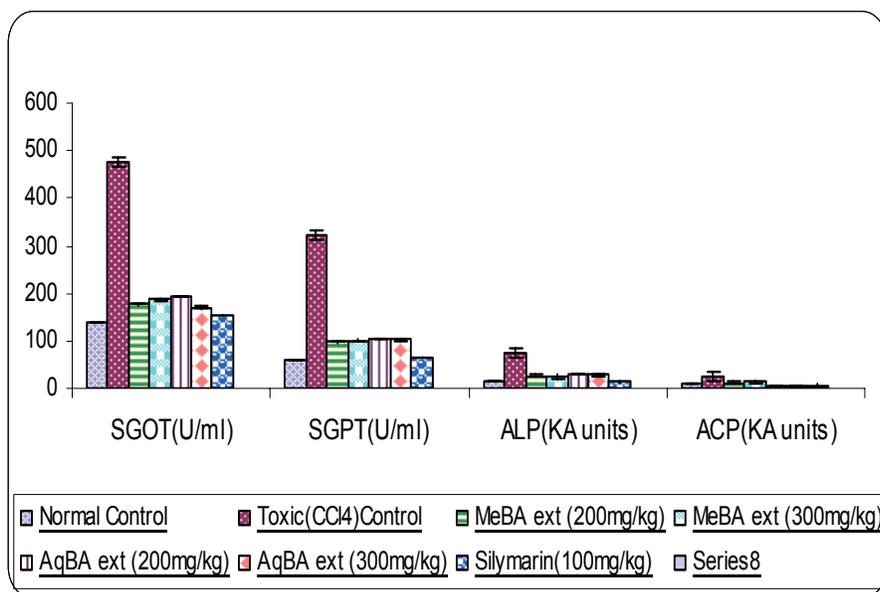


Fig. 1(a): Serum enzyme levels methanol extract treated animals

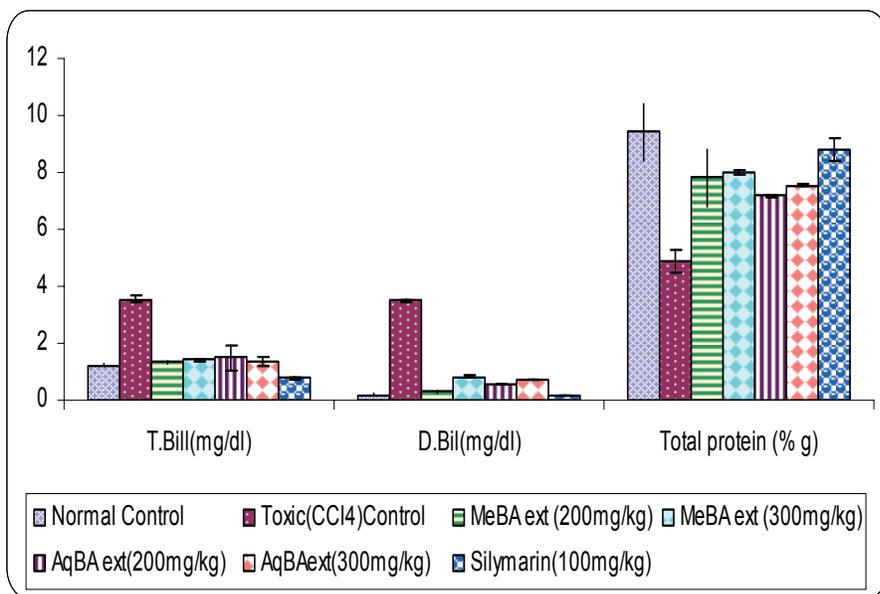


Fig. 1(b): Serum enzyme levels methanol extract treated animals

**Table 2: Effects of methanolic extract of arial parts *Berberis asiatica* on CCl<sub>4</sub> induced hepatotoxicity**

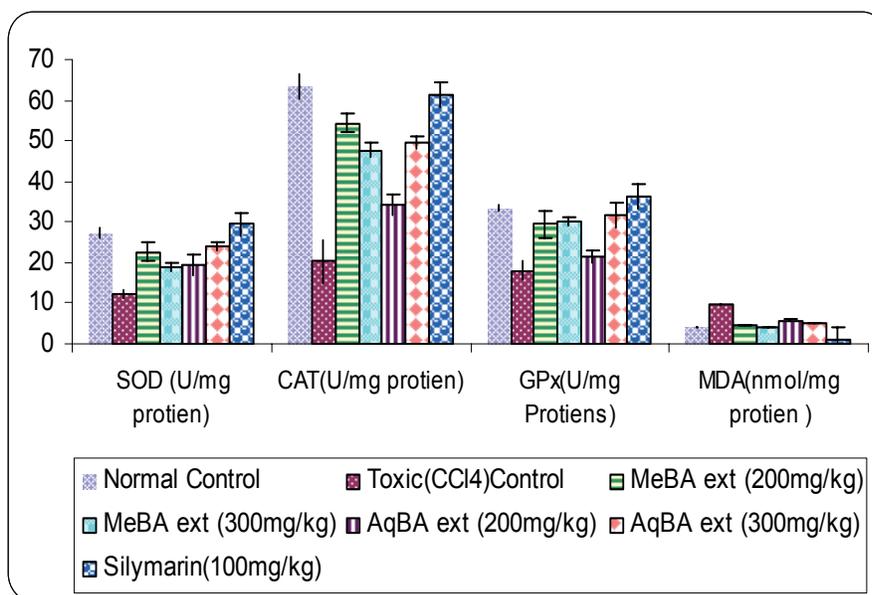
Group	SGOT (U/ml)	SGPT (U/ml)	ALP (KA units)	ACP (KA units)	S.bilirubin(SB)(mg/dl)		Total protein g/dl
					T-Bill	D-Bill	
Normal Control	137.53 ± 4.22	58.98±3.12	14.57±1.40	9.41± 0.44	1.23±0.03	0.20±0.01	9.42±1.02
Toxic(CCl <sub>4</sub> )Control	475.77 ± 10.08**	320.52± 12.03**	74.34 ± 9.98**	26.41±0.83**	3.55±0.11**	1.50±0.03**	4.91±0.41**
CCl <sub>4</sub> +MeBA ext (200mg/kg) (%protection)	177.01± 3.11** ( 88.32)	97.14 ± 1.74** (85.40)	26.42 ± 1.64** (80.13)	11.98±0.22** (84.68)	1.26±0.02** (98.70)	0.19 ± 0.02** (96.90)	8.01±1.02** ( 106..30)
CCl <sub>4</sub> +MeBA ext (300mg/kg) (%protection)	187.31 ± 2.78* (85.28)	101.14±2.64* (83.88)	25.44 ± 3.40* (81.77)	12.41±1.42* (82.35)	1.45± 0.01* (90.31)	0.32 ± 0.04* (90.76)	7.81±0.11* (64.30)
CCl <sub>4</sub> + AqBa ext (200mg/kg) (%Protection)	184.11± 2.81* (86.34)	104.14± 1.32** (82.73)	32.15 ± 1.43** (70.55)	15.07±1.55** (66.70)	1.51± 0.44** (87.93)	0.67± 0.02** (63.00)	7.19± 0.04** (50.55)
CCl <sub>4</sub> +AqBa ext (300mg/kg) (%Protection)	170.20 ± 1.71** (90.34)	101.14±1.67** (83.88)	29.42 ± 2.00** (75.11)	14.51± 1.10** (70.0)	1.61± 0.19** (83.62)	0.75± 0.01** (57.0)	7.55± 0.06** (58.53)
CCl <sub>4</sub> +Silymarin (100mg/kg) (%protection)	151.94 ± 2.47** (95.73)	65.45± 1.13** (97.52)	12.41± 1.23** (103.5)	7.34± 0.41** (112.17)	0.79±0.04** (118.96)	0.24 ± 0.01** (100.7)	8.81±0.41** (134.0)

Value are mean ± SD (n=6), \*Statistically different at p<0.05, \*\* Statistically different at p<0.001

**Table 3:Effects of Methanol extracts of *Berberis asiatica* aerial on Liver antioxidant enzymes and Lipid peroxidation on CCl<sub>4</sub> induced hepatotoxiciy**

Group	SOD (U/mg protein)	CAT (U/mg protein)	GPx (U/mg protiens)	MDA (nmol/mg protien )
Normal Control	27.20 ± 1.26	63.32 ± 3.10	33.32 ± 0.48	3.95± 0.11
Toxic(CCl <sub>4</sub> )Control	12.33 ± 1.04*	23.55 ± 5.45*	18.10 ± 2.36*	9.88± 0.03*
CCl <sub>4</sub> +MeSI ext (200mg/kg)	24.90 ± 1.56**	52.30 ± 4.03**	28.89 ± 3.61**	4.51. ± 0.02**
CCl <sub>4</sub> +MeSI ext (300mg/kg)	21.48 ± 3.83*	51.18 ± 2.01*	28.15 ± 1.45*	4.36 ± 0.02*
AQBA ext (200mg/kg)	22.68 ± 2.13**	48.31 ± 2.02**	29.51 ± 3.30**	4.65. ± 0.07**
AQBA ext (300mg/kg)	23.48 ± 1.02*	47.61 ± 2.01*	30.05 ± 1.04*	4.26 ± 0.03*
CCl <sub>4</sub> +Silymarin(100mg/kg)	19.40 ± 1.01**	61.14 ± 2.47**	36.14 ± 4.24**	1.01± 0.05**

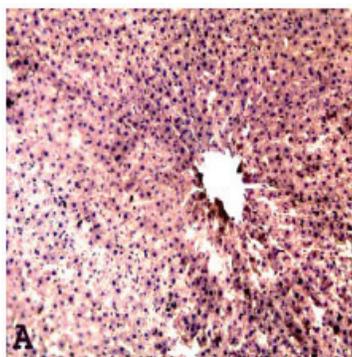
Value are mean ± SD (n=6), \*Statistically different at p<0.05\*\*,Statistically different at p<0.001



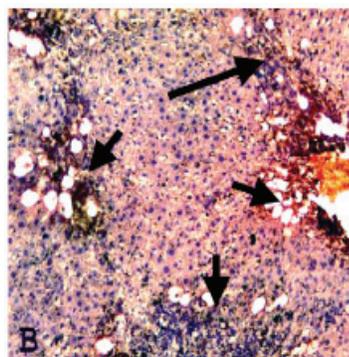
**Fig. 2: Antioxidant effect of methanol extract**

As given in table (3) reduced activities of enzymic and non-enzymic antioxidants and enhanced activity of lipid peroxidation were seen in the CCl<sub>4</sub> –treated group, where as standard Silymarin and the drug

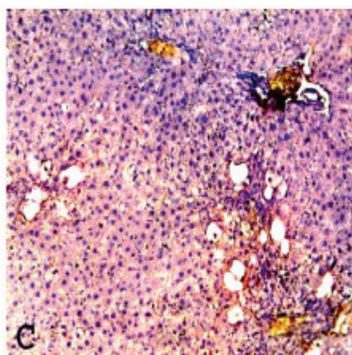
treated groups showed significant (p< 0.001) rise in antioxidant levels with reduction in lipid peroxidation level when compared with the CCl<sub>4</sub> –treated control group(fig 2)



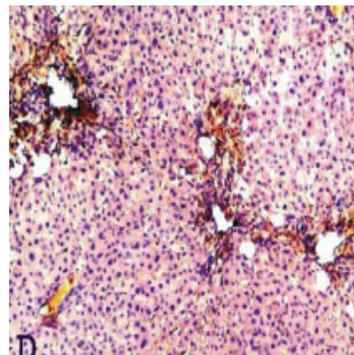
**Fig. A: Liver cells of normal rats**



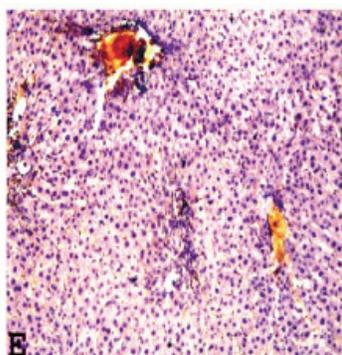
**Fig. B: Liver cells of rats intoxicated with CCl<sub>4</sub>**



**Fig. C: MeBA (200mg/kg) treated Liver cells**



**Fig. D: AqBA (300mg/kg) treated Liver cells and intoxicated with CCl<sub>4</sub> and intoxicated with CCl<sub>4</sub>**



**Fig. E: Silymarin treated (100mg/kg) treated Liver cells and intoxicated with CCl<sub>4</sub>**

From the tables and figures it is clear that the both MeBA and AqBA showed dose dependent hepatoprotective activity. However MeBA (200mg/kg) exhibited relatively higher protective action which is comparable with standard drug, silymarin (100mg/kg).

The Histopathological studies of liver showed fatty changes, swelling and necrosis with the loss of hepatocytes in CCl<sub>4</sub>-treated group, whereas the drug treated groups showed absence of cell necrosis, but with minimal inflammatory conditions around the central vein. The MeBA (200mg/kg, p.o.) treated group showed minimal inflammatory condition with near normal liver architecture. The silymarin treated group

showed almost normalization of fatty accumulation and necrosis. (Fig, A-E).

#### DISCUSSION

CCl<sub>4</sub>- is biotransformed by cytochrome P-450 in the liver endoplasmic reticulum to the highly reactive trichloromethyl free radical. This free radical in turn reacts with oxygen to form trichloromethylperoxy radical, which may attack lipids on the membrane of endoplasmic reticulum more readily than the trichloromethyl free radical. The trichloromethyl peroxy radical leads to elicit lipid peroxidation, the distribution of Ca<sup>+</sup> homeostasis, elevation of hepatic enzymes and finally results in cell death<sup>25</sup>.

It can be concluded from this investigation that, among the aqueous and methanolic extracts tested, the methanolic extract (200mg/kg) of the roots of *Berberis asiatica* possess more effective hepatoprotective activity against CCl<sub>4</sub> intoxication in rats because of its antioxidant bearing capacity. Our further detailed studies may, however, confirm the utility profile of this drug.

#### ACKNOWLEDGEMENT

The author is thankful to Dr. H.B Singh of National Institute of Science communication and Information recourses (NISCAIR), New Delhi, India for identification and authentication of plants

#### REFERENCE

- Hemnani T, Parihar M S, Reactive Oxygen Species and Oxidative DNA Damage. *Indian J. Physiol. Pharmacol.*, 1998; 42: 440-4.
- Yoshikawa T., Tanaka H, Yoshida N, Seto O, Sugino N. and Kondo, M. Adjuant arthritis and Lipid Peroxide Protection by Superoxide Dismutase. *Lipid Peroxide Res* 1983; 7, 108-110.
- Halliwell B. and Gutteridge J.M.C. Lipid Peroxidation, Oxygen Radicals; Cell Damage and Antioxidant Therapy. *Lancet* 1984; 1: 1396-1397.
- Yuda, Y., Tanaka, J., Hirano, F., Igarani, K. and Snatch, T.J. Participation of Lipid Peroxidation. In Rat Pertusis Vaccine Pleurisy. *Chem. Pharm. Bulletin* 1991; 39, 505-506.
- Brattin W J, and Glenda E A. Pathological Mechanisms in Carbon Tetrachloride Hepatotoxicity. *J. Free Radical Biol. Med.* 1985; 1: 27-38.
- Thresiamma K C, Kuttan R. Inhibition of liver fibrosis by ellagic acid. *Indian Journal of Physiology and pharmacology.* 1996; 40 [4]: 363-366
- Osawa T, Kavakishi S, Namiki M, Kuroda Y, Shankal D M, and Waters M.D. Role of dietary antioxidants in protection against oxidative damage. In 'Basic Life Sciences: Antimutagenesis and Carcinogenesis Mechanisms II. Mechanism II' Vol. (Eds Kuroda Y, Shankel DM and Waters MD. Plenum Press), New York, pp. 1990; 52: 139-153.
- Subramanian A, Evans, D A, Rajasekharan S, Pushpangadan P. Hepatoprotective Activity of *Trichopus Zeylanicum* Extract Against Paracetamol-Induced Hepatic Damage in Rats. *Indian J. Exp. Biol.* 1998; 36: 385-389.
- Watt G, *Economic product of India V.* The superintendent of government of printing, India, 1883.
- Kirtikar K R, Basu B D. *Indian Medicinal Plants*, I. Allahabad Latit Mohan Basu and Co 1933.
- Anonymous. The wealth of India *Berberis* Linn. (Berberidaceae). New Delhi. Ambastha SP publications and information directorate. CSIR, 1988; pp 114-116.
- Bhakuni D S, Shoeb A, Popli S P. Studies on Medicinal Plants, part I. Chemical Constituent of *Berberis asiatica* Roxb. *Indian Journal of chemistry*, 196; 6: 123.
- Rastogi R.P, Mehrotra B N. *Compendium of Indian Medicinal Plants.* 1993; Vol 3: New Delhi: Publication and Information Directorate.
- Shah N C, Joshi M C. An ethnomedicinal study of Kumoun region of India. *Economic Botany* 1971; 25: 411-422.
- Peach K, Tracey M V. Springer-Verlag, Heidelberg 1955; 4: 142 - 9.
- Tiwari B K, Khosa R L. Evaluation of hepatoprotective activity of *Sphaeranthus indicus* flower heads. *Journal of Natural Remedies*, (2008; 2(8): 173-178.
- Lin C C, Yen M H, Lo T S, Lin J M, Evaluation of the hepatoprotective and antioxidant activity of *Boehmeria nivea* var. *nivea* and *B. nivea* var. *tenacissima*, *Journal of Ethnopharmacology* 1998; 60: 9-17
- King J. The transferase Alanine and aspartate transaminase. In Van, D(Ed). *Practical clinical enzymology.* No strand Co. Ltd London, (1965b) pp 121-138.
- King J. The hydrolases acid and alkaline phosphatase In Van, D(Ed). *Practical clinical enzymology.* No strand Co. Ltd London, (1965a) pp 208.
- Malloy H T, Evelyn K A, The determination of Bilirubin with photoelectric colorimeter. *Journal of Biological chemistry* 1937; 119: 481-490.
- Marklund S L, Marklund G. Involvement of Superoxide anion radical in auto-oxidation of pyrogallol and convenient assay for superoxide dismutase. *European journal of Biochemistry* 1974; 47: 469-474.
- Rotruck J T, pope A L, Ganther H E. Selenium Biochemical role of a component of glutathione peroxidase. *Purification and assay of science* 1973; 179:588-590.
- Aebi H. Catalase in vitro, *Methods Enzymol.* **105** (1984) 121-126.
- Ohkawa H, Ohisi N, Yagi K. Assay of lipid peroxidase in animal tissue by thiobarbituric reaction. *Anal Biochem* 1979; 95:351-358.
- Clawson GA. Mechanism of carbon tetrachloride hepatotoxicity. *Pathology and immunopathology Research* 1989; 8:104-112