



CARBOXYMETHYL ETHERS OF LOCUST BEAN GUM- A REVIEW

PARAMITA DEY^{1*}, BISWANATH SA¹ AND SABYASACHI MAITI²¹Department of Pharmaceutical Technology, Jadavpur University, Kolkata-700032, West Bengal, India, ²Gupta College of Technological Sciences, Ashram More, G.T. Road, Asansol-713301, West Bengal, India, Email: paramita.dey6@gmail.com

Received: 11 Dec 2010, Revised and Accepted: 15 Jan 2011

ABSTRACT

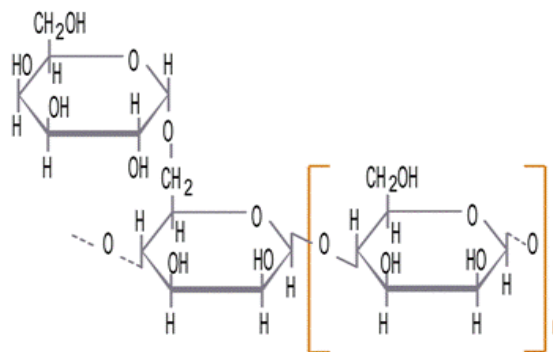
Locust bean gum is a popular natural polymer which is mostly used in food industry as well as in pharmaceutical industry. This natural polymer is conventionally used as an excipient in manufacturing different formulations which mainly depends on its thickening and gelling property. Locust bean gum can easily be modified to its carboxymethyl derivative which in turn can be used as sustained release delivery carrier as rate controlling polymer. The modified gum shows improved physical properties like good aqueous solubility, acceptable solution viscosity and clarity compared to the native one. The main reaction is carried out in presence of mono-chloroacetic acid under catalytic conditions like sodium hydroxide in aqueous environment and sodium bicarbonate in non-aqueous environment. But the degree of substitution of carboxy methyl group in the structure of locust bean gum is an important criterion. Various scientists have put forward various sophisticated methods to determine the degree of substitution. The most classical method is done by titrimetric method. Various other modern instruments are coming up for this purpose like Fourier Transform Infrared Spectroscopy method, Nuclear Magnetic Resonance method and Raman Spectroscopy method. This review describes the different methods for the preparation of carboxymethyl locust bean gum and the analysis of the degree of substitution of carboxymethyl group by using different analytical techniques.

Keywords: Locust Bean Gum, Carboxymethylation, Degree of Substitution, Analytical methods

INTRODUCTION

Polymers are macromolecules comprised of repeating units of small molecules, the monomer. The monomers can be linked together to form linear polymer or branched polymer or cross linked polymers. Linear polymer and branched polymer are referred to as thermoplastic materials as they flow on heating. They also show solubility in certain solvents. Locust bean gum is a branched polymer. Biopolymers or natural polymers are an attractive class of biodegradable polymers since they are derived from natural sources, easily available, relatively cheap and can be modified by suitable reagent. The specific application of plant-derived polymers in pharmaceutical formulations include their use in the manufacture of solid monolithic matrix systems, implants, films, beads, micro particles, nano particles, inhalable and injectable systems as well as viscous liquid formulations¹⁻³. The successful formulation of a stable and effective dosage form therefore depends on the careful selection of excipients. The present trend focuses on an increasing interest in the use of natural ingredients in food, drugs and cosmetics⁴⁻⁸.

Locust bean gum, a non starch polysaccharide consisting of galactose and mannose in the ratio 1:4 and hence they are known as galactomanan⁹. The mannose elements form a linear chain linked with galactopyranosyl residues as side chain at varying distances depending on the plant origin¹⁰. Being a galactomanan locust Bean Gum has a wide application in pharmaceutical field. It is also known as Carob bean gum and is derived from the seeds of the leguminous plant *Ceratonia siliqua* Linn belonging to the family Fabaceae. This gum is widely cultivated in the Mediterranean region and to smaller extent also in California. The brown pods or beans of the locust bean tree are processed by milling the endosperms to form locust bean gum¹¹. It consists mainly of a neutral galactomanan polymer made up of 1, 4-linked D-mannopyranosyl units and every fourth or fifth chain unit is substituted on C6 with a D-galactopyranosyl unit¹². The ratio of D-galactose to D-mannose differs and this is believed to be due to the varying origins of the gum materials and growth conditions of the plant during production. The physico-chemical properties of galactomanan are strongly influenced by the galactose content¹⁴ and the distribution of the galactose units along the main chain¹⁵. Longer galactose side chains produce stronger synergistic interactions with other polymers¹⁴ and greater functionality¹⁵. Since it is a neutral polymer and its viscosity and solubility are therefore little affected by pH changes within the range of 3-11^{16,17}.

**Fig. 1: Structure of locust bean gum¹⁶**

Various properties are there which make locust bean gum a good choice in drug delivery.

- They are biocompatible, biosorbable and biodegradable in nature.
- It is non-teratogenic and non-mutagenic according to Joint FAO/WHO Expert Committee on Food Additives held in Geneva, April'75.
- Acceptable shelf-life.
- Degradation products are excreted readily.

Nowadays a trend has come to modify non-starch polymer in order to modify their physico-chemical properties. Thus the modified natural polymers can be defined as the natural polymers altered to improve their biodegradation profile and also physico-chemical characteristics. Generally labile polar functionalities are added to the polymer to enhance the degradability of the polymer. The extent and nature of polymer modification is vital as excess modification can hamper the biodegradation and the added functional group may be converted to toxic degradation products¹⁸⁻²¹. This modification of natural polymers is achieved by chemical modification or enzymatic alteration. The chemical modification involves harsh conditions in comparison to the enzymatic method.

Locust bean gum, non-starch polysaccharides can be chemically modified to alter their physico-chemical properties to improve

specific functional properties. Carboxymethylation is a well-known etherification process with carboxymethyl groups²². Carboxymethylation of polysaccharides is a widely studied conversion since it is simple and leads to products with a variety of promising properties. Carboxymethyl cellulose exhibits improved solubility and has been used in the food, cosmetic, detergent, paper, pharmaceutical and textile industry²³. Carboxymethyl guar gum, locust bean gum and xanthan gum have been used in transdermal drug delivery system²⁴. Other chemical modifications of polysaccharides are also carried out by esterification, oxidation and hydroxypropylation²⁵. Carboxymethylation generally increases the hydrophilicity and solution clarity of the polysaccharides and makes it better soluble in aqueous system¹. This high soluble polymer is a good candidate for preparation of different novel drug delivery like beads, microparticles or nanoparticles. The objective of this review article is to focus on the different methods of synthesis of carboxymethylated ether of Locust bean gum and its determination of the degree of substitution by using different methods.

SYNTHESIS OF CARBOXYMETHYLATED LOCUST BEAN GUM

Carboxymethylated Locust bean gum can be synthesized by either of the two methods given by different scientists

- By following non-aqueous method⁹.
- By following aqueous method²⁶.

Synthesis by non-aqueous method

Finely powdered Locust bean gum and sodium bicarbonate are mixed well manually in a pestle mortar. To this 1% v/v of ethanol and solid monochloro-acetic acid is added. The reaction was carried out at ambient or elevated temperature (60, 80, 98°C) for 2 hours with intermittent manual mixing with a glass rod, by addition of dilute acetic acid to phenolphthalein end point. Salts formed were removed by repeated washings with 70% aqueous ethanol for 15 minutes followed by 100% ethanol and solvent exchange drying²⁷. The derivatized gum was tested for sodium using uranyl magnesium acetate spot test as well as atomic absorption spectroscopy, were found to be free from sodium ions, by these washings²⁸. A number of carboxymethyl derivatives of differing degree of substitution values were prepared by varying the ratio of catalyst and the reagent. In a similar way Carboxymethylation was carried out without prior surface wetting with alcohol and the derivitized dry products were recovered. Various studies of carboxymethylation of Locust bean gum were carried out to optimize the reaction condition to increase the product yield, reaction efficiency and the degree of substitution^{29,30}.

Synthesis by aqueous method

Locust bean gum was derivitized to sodium carboxymethyl Locust bean gum by mixing it with 4ml water heated to 80°C for 15 minutes and cooled. Then 56% w/v of ice-cold sodium hydroxide solution was added drop wise over a period of 45 minutes. Monochloroacetic acid solution was added slowly for a period of 1 hour to the above mixture and maintained at 15°C. The temperature of the mixture was raised slowly to 65°C and stirred for another 1 hour. The wetted mass was washed with methanol for 15 minutes. The pH of the suspension was adjusted to neutrality with glacial acetic acid. Then it is dried at 50-60°C³⁰.

Analytical tools for the structure characterization of carboxymethylated locust bean gum

The total degree of substitution can be defined as the average number of functional groups introduced in the polymer. This mainly determines the properties of polysaccharides derivatives including carboxymethylated products. The functionalization even affect the properties of that particular polymer^{31,32}. In most cases it is found that partially substituted polysaccharide is much more potent compared to fully substituted ones. Since fully substituted polysaccharides may undergo undesirable reactions like oxidation or depolymerization³³. The exact determination of degree of substitution and functionalization pattern is important for the optimization of reaction conditions in order to understand the structure property relationship. The degree of substitution should

be monitored to optimize the properties of the modified polysaccharides for specific manufacturing process. Carboxymethylation to low extent may lead to an uneven distribution of carboxymethyl moieties. Various methods are there to analyze the total degree of substitution. A brief account of this is furnished in this review article.

Determination of degree of substitution by wet method

The classical methods to determine the degree of substitution is by simple acid-base titration. This method is very useful because no expensive equipments are required and at the same time degree of substitution value is also very reproducible provided the procedure is carried out very carefully³⁴. Here the conversion of the salt form to free acid form and *vice versa* was carried out. This can be done by taking the acid form of the carboxymethylated polysaccharide, which can be obtained by a treatment of the sodium salt form of the polymer dispersed in ethanol with concentrated hydrogen chloride, can be titrated with a sodium hydroxide solution of known molarity³⁵. Mostly applied is the back titration method, which was proposed as standard procedure of CMS³⁵ and CMC³⁶. The sodium salt of the polymer is converted to the free acid form. Subsequently, aqueous sodium hydroxide is added to a known amount of the free acid form leading to the sodium carboxylate. The excess of sodium hydroxide is back titrated permitting to degree of substitution³⁵⁻³⁷. According to Mukhopadhyay et al. degree of substitution is calculated by³⁸;

$$DS = \frac{0.162 A}{1 - 0.058 A}$$

DS= Degree of substitution

A= Milliequivalents of NaOH required per gram of sample

Determination of degree of substitution by fourier transform infrared spectroscopy method

The degree of substitution of carboxymethylation was quantitatively determined by Fourier Transform Infrared Spectroscopy by calculating the ratio between the intensity of hydrogen bond to that of intensity of carboxyl stretching of ether. Both the native and modified gum samples were blended with solid potassium bromide and the pellet was prepared. The spectra were scanned from 400 to 4000 cm⁻¹ under dry air at room temperature¹. Infrared measurement was carried out in the transmission mode in which the infrared beam directly passes through the sample and spectral data were then converted from transmittance into absorbance unit¹⁰. Fourier Transform Infrared Spectra were plotted as relative intensity against wave number cm⁻¹. Band area changes with increase in degree of substitution³⁹⁻⁴¹. Fourier Transform Infrared Spectra indicated the introduction of carboxymethyl moiety whose intensity increased with increase in degree of substitution.

Determination of degree of substitution by raman spectroscopy

Raman Spectroscopy was first applied to determine the degree of acetylation and succinylation in starch⁴². Raman Spectroscopic methods have also been successfully adopted in the quantitative analysis of chemically modified food proteins⁴³. However, for the derivative of non-starch polysaccharides Raman Spectroscopy has only been used to identify new or substituted groups and to monitor reaction kinetics. An analytical technique based on Raman Spectroscopic methods was developed with an aim of replacing traditional wet chemistry method.

Raman Spectra of native and modified locust bean gum were recorded using a Renishaw Raman Imaging Microscope. Solid samples were prepared on microscopic slides. Raman shift was calibrated by a silicon slide. The spectra were collected at room temperature in the dark 30 seconds exposure time with 2cm⁻¹ resolution. Generally 10-20 scans were run. Raman spectra are collected in triplicate and plotted as relative intensity against Raman shift in wave-number (cm⁻¹)

Determination of degree of substitution by nuclear magnetic resonance method

At present, the most important and sophisticated instrument used for measurement of degree of substitution is ^{13}C CP/MAS NMR and solution ^{13}C NMR spectroscopy. From the ^{13}C CP/MAS NMR spectrum the average degree of substitution can be calculated from the ratio of area of the carboxyl signals. Compared with ^{13}C CP/MAS NMR spectra of dissolved samples are significantly increased which enables spectra recorded in this way to be evaluated for the purpose of determining the total degree of substitution⁴⁴.

Properties

One of the most important goals of carboxymethylation of locust bean gum is to obtain water soluble derivatives. This modified gum can be used typically in pharmaceutical purposes as rate controlling polymer. They behave as a typical polyelectrolyte.

The interaction of carboxylic groups with multivalent metal cations can be used to form so called ionotropic gels which are predominantly stabilized by the electrostatic interactions. In addition interactions between the OH groups of the polymers and the metal ions contribute to the stability and water insolubility of these polymeric aggregates. This modified gum thus has the capacity to the formation of gelled beads through ionotropic gelation with trivalent ions^{26, 45, 46}. Maiti et al. found that Glipizide, oral anti-diabetic drug can be loaded in the modified polymer beads which is useful for oral drug delivery. The drug release from the beads was prolonged upto 10 hour in alkaline dissolution medium depending upon the concentration of gelling agent (1-5%)⁴⁷.

CONCLUSION

Thus a non-starch polysaccharides locust bean gum which is a galactomannan can easily be carboxymethylated in order to modify their physical characteristics like solubility, solution viscosity and clarity. It can be modified by both aqueous and non-aqueous method too. In both the cases, carboxymethylation is simple, cost effective and eco-friendly. Thus the carboxymethylated locust bean gum developed by these methods show good degree of substitution. This modified gum finds use in the preparation of microparticle and other sustained release preparation. Their degree of substitution has been determined by traditional wet chemistry method and also by applying modern analytical techniques like FTIR, Raman Spectroscopy and NMR Study. When compared to the traditional wet chemistry methods, the spectroscopic methods developed in this study are relatively simple, fast without the use of toxic chemicals requiring smaller sample size. Thus these techniques are superior as they give both qualitative and quantitative measurement.

REFERENCES

- Pandey R, Khuller G. Polymer based drug delivery systems for mycobacterial infections. *Curr Drug Deliv* 2004; 1:195-201.
- Chamarthy SP, Pinal R. Plasticizer concentration and the performance of a diffusion controlled polymeric drug delivery system. *Colloids Surf. and Physicochem. Eng. Asp* 2008; 331:25-30.
- Alonso SM, Teijeiro D, Remunan LC, Alonso MJ. Glucomanan, a promising polysaccharide for biopharmaceutical purposes. *Eur J Pharm Biopharm* 2008; 72:453-462.
- Tonnesen HH, Karlsen J. Alginate in drug delivery system. *Drug Deliv and Indus Pharm* 2002; 28(6): 621-630.
- Miyazaki S, Kubo W, Attwood DJ. Oral sustained delivery of Theophylline using in-situ gelation of sodium alginate. *J Cont. Rel* 2000; 67(2): 275-280.
- Bhardwaj TR, Kanwar M, Lal R, Gupta A. Natural Gums and modified natural gums as sustained release carriers. *Drug Deliv and Indus Pharm* 2000; 26:1025-1038.
- Sultzbaugh KJ, Speaker TJJ. A method to attach lectins to the surface of spermine alginate microparticles based on the avidin biotin interaction. *J of Microencap* 1996; 13(4):363-375.
- Quong D, Neufeld RJ. DNA encapsulation within co-guanidine membrane coated alginate beads and protection from extracapsular nuclease. *J of Microencap* 1999; 16(5):573-585.
- Parvathy KS, Susheelamma NS, Tharanathan RN, Gaonkar AK. A simple non-aqueous method for carboxymethylation of galactomannans. *Carbohydr. Polym* 2005; 62:137-141.
- Sharma BR, Dhuldhoya NC, Merchant SN. Glimpses of Galactomannans. *Sci Tech Entrepreneur* 2008; 3: 1-10.
- Beneke CE, Viljeon AM, Hamman JH. Polymeric plant derived excipients in drug delivery. *Molecule* 2009; 14:2602-2620.
- Venkataramu MP, Gowda DV, Rajesh KS, Shiva KHG. Xanthan and locust bean gum (from *Ceretonia siliqua*) matrix tablets for oral controlled delivery of propranolol hydrochloride. *Asian J Pharm Sci* 2007; 2(6): 239-248.
- Dea ICM, Morrison A. Chemistry and interactions of seed galactomannans. *Adv Carbo Chem and Biochem* 1975; 31: 242-312.
- ER Morris. Mixed polymer gels. London (England): Elsevier Applied Science; 1990.
- Launay B, Doublier JR, Cuvelier G. Flow properties of aqueous solutions and dispersions of polysaccharides. London (England): Elsevier Applied Science; 1986.
- Deuel H, Neukom H. Some properties of Locust Bean Gum. *Advances in Chem* 1954; 11: 51-61.
- Yang H, Zhou SB, Deng J. Preparation and properties of hydrophilic-hydrophobic chitosan derivatives. *J. Appl. Polym. Sci* 2004; 92(3):1625-1632.
- Cao Z, Ge H, Lei S. Studies on synthesis and adsorption properties of chitosan cross-linked by glutaraldehyde and Cu(II) as template under microwave irradiation. *Eur. Polym. J* 2001; 37(10):2141-2143.
- Chen XG, Park HJ. Chemical characteristics of O-carboxymethyl chitosans related to the preparation conditions. *Carbohydr Polym* 2003; 53(4):355-359.
- Rhazi M, Desbrieres A, Tolaimate M, Rinaudo P. Influence of the nature of the metal ions on the complexation with chitosan.: Application to the treatment of liquid waste. *Eur. Polym. J* 2002; 38(8):1523-1530.
- Wang XH, Du YM, Liu H. Preparation, characterization and antimicrobial activity of chitosan-Zn complex. *Carbohydr Polym* 2004; 56(1):2-26.
- Yuen SN, Choi SM, Phillips DL, Ma C. Raman and FTIR spectroscopic study of carboxymethylated non-starch polysaccharides. *Food chem.* 2009; 114:1091-1098.
- Togrul H, Arslan N. Production of carboxymethyl cellulose from sugar beet pulp cellulose and rheological behavior of carboxymethyl cellulose. *Carbohydr Polym* 2003; 54:74-84.
- Murthy SN, Hiremath SRR, Paranjothy KKK. Evaluation of carboxymethyl guar films for the formulation of transdermal therapeutic systems. *Int J Pharmaceu* 2004; 272:11-18.
- Sierakowski MR, Mitas M, Desbrieres J, Rinacedo M. Specific modifications of galactomannans. *Carbohydr Polym* 2000; 42:51-57.
- Maiti S, Dey P, Banik A, Sa B, Ray S, Kaity S. Tailoring of locust bean gum and development of hydrogel beads for controlled oral delivery of glipizide. *Drug Deliv* 2010; 17(5): 288-300.
- Green JW. O-carboxymethyl cellulose. *Methods in Carbohydr Chem* 1963; 3:325-326.
- Vogel AI. Text book of macro and semi micro qualitative inorganic analysis. 4th ed. London (England): Longman Publisher; 1989.
- Khalil MI, Beliakova MK, Aly AA. Preparation of some starch ethers using semi-dry process. *Carbohydr Polym* 2001; 46:217-226.
- Khalil MI, Hasehem A, Habiesh A. Carboxymethylation of maize starch. *Starch/Starke* 1990; 42:60-63.
- Shogren RL. Starch: properties and material applications in biopolymers from renewable resources. 1st ed. Berlin: Springer Publisher; 1998.
- Phillips GO, Williams PA. Handbook of hydrocolloids. 1st ed. Boca Raton. FL (USA): CRC Press; 2000.
- Yalpani M. Polysaccharides: Synthesis, modifications and structure/property relations. New York: Elsevier; 1988.
- Eyler RW, Klug ED, Diephuis F. Determination of degree of substitution of sodium carboxymethylcellulose. *Analytical Chem* 1947; 19:24-27.
- Olaf W, Petra M. Determination of the substitution pattern of cationic starch ethers. *Starch/Starke* 1997; 49(11):453-458.

36. Miller GL. Determination of reducing sugar by DNS method. *Analytical Chemistry* 1959; 31:426-428.
37. Welcher FJ. *Standard methods of chemical analysis*. 6th Ed. New York: Krieger Publisher; 1975.
38. Mukhopadhyaya S, Mitra BC, Palit SR. Determination of degree of substitution in sodium carboxymethyl cellulose. *Analy Chem* 1975; 45 (9):1775-1777.
39. Mathlouthi M, Koenig JL. Vibrational spectra of carbohydrates. *Adv in Carbohydr chem and biochem* 1986; 47:75-89.
40. Zbankor RG. *Infrared spectra of cellulose and its derivatives*. New York: Consultant Bureau; 1966.
41. Zbankor RG, Andrianov VM, Marchewka MK. Fourier transform Infrared spectroscopy and Raman Spectroscopy and structure of carbohydrates. *J of Molecul Struc* 1997; 637-654.
42. Phillips DL, Xing J, Liu H, Chong CK, Corke H. General application of Raman spectroscopy for the determination of level of acetylation in modified starches. *Cereal Chem* 1999; 76:439-443.
43. Wong HW, Phillips DI, Ma C. Raman spectroscopic study of aminated food proteins. *Food Chem* 2007; 105:784-792.
44. Atalla RH, Gast JC, Sindorf DW, Bartuska VJ, Maciel GE. ¹³NMR spectra of cellulose polymorphs. *J of Am Chem. Society* 1989; 10:3249-3251.
45. Murata Y, Malda T, Miyanioto E, Kawashima S. Natural gums and modified natural gums as sustained release carriers. *Int. J. Pharm* 1993; 96:139-145.
46. Ray S, Maiti S, Sa B. Preliminary investigation on the development of Diltiazem Resin complex loaded carboxymethyl xantahn beads. *AAPS Pharm Sci Tech* 2008; 9:295-301.
47. Halder A, Mukherjee S, Sa B. Development and evaluation of polyethyleneimine-treated calcium alginate beads for sustained release of Diltiazem. *J. Microencap* 2005; 22:67-80