

## ROLE OF CADHERIN SWITCHING IN EMT AND PROSTATE CANCER METASTASIS - A TOPIC REVISITED

SURESH P.K.<sup>1</sup> & NATHAWAT L.<sup>2</sup>

<sup>1</sup>School of Bio Sciences & Technology, VIT University, Vellore Dt. PIN:632014, <sup>2</sup>M.Sc. Biomedical Genetics, VIT University, Vellore, Vellore Dt. Pin:632014. Email: p.k.suresh@vit.ac.in;indian.ethos@gmail.com

Received: 21 Jan 2014, Revised and Accepted: 21 Apr 2014

### ABSTRACT

The conversion of the sessile epithelial cells to the motile mesenchymal phenotype (EMT transition) involves the characteristic switching of E-cadherin to N-cadherin and is a "signature-like event", involving the TGFβ1-mediated pathway, in the process of invasion and metastasis of prostate cancer cells – a process commonly observed in other cancer cells. The transcriptional epigenetic repression of E-cadherin is associated with and regulated by the expression of ZEB1 – zinc finger homeo-domain transcription repressor, which in turn, is regulated by specific microRNAs. The role of IGF-1, correlatable with ZEB1, through equivocal, may have an important staging-dependent differential role in prostate cancer. Other transcription factors (Snail, Slug & E-47), when expressed, induce, among other signaling molecules, the expression of IGF-1 and Wnt-5. This, in turn, causes E-cadherin repression. One of the major, common downstream pro-survival effector protein is PI3K/Akt and is regulated both by the Ras as well as the TGF-β1 pathways. This pivotal protein is known to protect the cells against TGFβ1-mediated apoptosis and plays an important role in EMT and metastasis. Repression of E-cadherin, is accompanied by the Twist1-dependent expression of N-cadherin. Corroborative evidence supports the abnormal activation of the Wnt/β catenin pathway and this pathway has been strongly implicated in prostate cancer invasion and metastasis pathways while catenin-independent pathways have also been reported apart from important epigenetic mechanisms regulating the inhibitors of the pathway like (Wnt inhibitory factor-1 -WIF-1). This review provides the reader with an update on the role of important signaling molecules and a better molecular understanding of cadherin switching – lessons that can be applied in cancer biology and chemoprevention by ethno-pharmacological and bio-pharmaceutical approaches.

**Keywords:** E-cadherin, N-cadherin, EMT, Transcription factors, Metastasis, Prostate cancer.

### INTRODUCTION

Prostate Cancer, more precisely prostate adeno-carcinoma, is one of the most commonly diagnosed cancers in men (globally), and one of the leading causes of cancer-related deaths in western countries with the incidence being lower in Asia. However, the changing lifestyle and the westernization in terms of increased consumption of fatty food and obesity, apart from improvements in diagnosis and life expectancy, has contributed to the rising incidence of prostate cancer in developing countries. Further, an unfavorable stage distribution has also been reported[1],[2][3] Since this disease is the outcome of a complex interplay between environmental factors as well as the underlying genotype, race and ethnicity are important considerations in the evaluation of gene-gene and gene-environment interactions[4]. This changing risk profile is also mirrored in the decrease in the risk between Asian immigrants and natives in the developed countries [3]. Deaths due to prostate cancer occur mostly due to metastases formed by the tumor cells at secondary sites, particularly bones. Prostate carcinoma forms metastases at secondary sites in a well recognised pattern which involve the axial skeleton and local lymph nodes[5][6]. Metastases are seen in other sites as well such as lungs, brain and liver, but to a lesser extent [6][7]. Skeletal metastases formed by prostate cancer are more frequently osteoblastic in nature and is known to follow an order in terms of frequency with differences in the stage-dependent distribution in the different regions of the spine. This prediction for the spinal localization, early in the metastatic process, is consistent with the reported backward spread via the veins in addition to the involvement of the vena cava-based process[8][9].

The propensity of prostate cancer cells to metastasize to secondary organs has been explained by a number of hypotheses. Batson proposed that retention of prostate cancer cells to bones might be due to the retrograde flow of prostate cancer cells in veins[10][11]. The famous "Seed and Soil" hypothesis for metastasis of cancer proposed by Paget suggests that the micro-environment of the secondary site (soil) determines the selectivity of the cancer cells (seed). This theory still holds forth today, as the potential of a tumor cell to metastasize to a secondary site is dependent on its

interactions with the micro-environment of secondary site as well as on factors, which promote tumor cell survival, angiogenesis, invasion and metastasis[12][13]. Also, there is evidence of prostate cancer cells in their journey towards acquiring the metastatic phenotype, become osteoblastic, through inductive influences with bone stromal cells. This leads to an alteration in critical transcription factors (Cbfa and MSX) which, in turn, can favor the expression of genes like osteopontin (OPN), osteocalcin (OC) and bone sialoprotein (BSP)[14][15]. There are certain common steps in the metastatic cascade that must occur in all forms of cancer. These sequential and selective steps, with certain stochastic components, involve the loss of cell adhesion at a primary site, invasion, migration, and survival and growth of tumor cells at a secondary site with heterogeneity, both within a single cell and between metastases, as one of its hallmark features[16][17]. The very first requirement for a cell to metastasize is that it loses its adhesion with surrounding cells. Loss of cell adhesion at a primary site is believed to be mediated by Epithelial-Mesenchymal Transition, where the epithelial cells lose their cell-cell junctions and characteristic features and become more motile [18]. In this regard, they re-active an embryonic program, wherein the sessile epithelial cells acquire mesenchymal features and become motile. Further, conversion renders the transformed cells with stem cell-like properties (resistance to therapy and apoptosis, apart from a decrease in senescence as well as the ability to evade the immune system) with the acquisition of the invasive phenotype being important for metastasis to occur[19],[20]. Cadherin switching is one of the aspect of epithelial-mesenchymal transition, where cells switch expression of their characteristic cadherins and express unusual cadherins at adherens junctions which affect the phenotype and behaviour of the cells due to a change in the isoform of the cell adhesion protein[21][22][23]. Loss of E-cadherin with increase in expression of N-cadherin is the most remarkable event occurring when a tumor cell acquires metastatic properties[24][25].

### Cadherins

Cadherins are the major cell adhesion molecules. They are calcium-dependent adhesion molecules and play a crucial role in the spatial

segregation of cell types and organisation of different tissues during embryonic development[26][27][28]. Cadherins interact with other cadherins on adjacent cells by a complex of proteins called catenins. The catenins bind to the actin cytoskeleton of the cell. The cadherin-catenin complex forms the classic adherens junctions which integrate the epithelial cells in a mechanical unit. Cadherins join cells together by homophilic binding, binds to the same type of cadherin on another cell. Cell adhesion by cadherins is mediated by both the homophilic binding of extra cellular domains and binding of cytoplasmic domain of cadherin with actin cytoskeleton[29][30]. Homophilic binding between the extracellular domains of cadherins is initiated and stabilized by binding of  $Ca^{2+}$ [31][32]. E-cadherin, also known as uvomorulin, is expressed on all the early embryonic cells of mammals. Later its expression is restricted to epithelial cells. Mesenchymal cells, which are less polarized and more motile than epithelial cells, express N-cadherin (neural cadherin) and various other cadherins such as R-cadherin and cadherin-11[33][34][35]. VE-cadherin is expressed specifically by endothelial cells at the junctional complex. Endothelial cells also express N-cadherin whose function is unknown as they are not expressed at the junctions. E-cadherin is expressed by epithelial cells where it provides the mechanical strength to the tissue, however many epithelium-derived cancer cells lose the expression of E-cadherin[36][37][38]. Enzymatic activity is not found in classical cadherins and catenins but in adherens junctions, they can associate with kinase and phosphatase enzymes such as Fer and PTP1B[39][40]. Adhesion of E-cadherin activates phosphatidylinositol 3-kinase (PI3-K) and Akt/protein kinase B[41]. Akt is a serine/threonine kinase which is activated by growth factors and integrin adhesion. Akt plays a regulating role in various metabolic pathways and apoptotic pathways. Phosphorylation of threonine 308 (Thr-308) and serine 453 (Ser-473) by 3-phosphoinositide dependent kinase 1 or phosphoinositide dependent kinase 2 results in activation of Akt[42]. Upon activation, Akt phosphorylates various substrates that suppress apoptosis. When a cell receives apoptotic signals, cell fate is determined by the balance between pro-apoptotic and anti-apoptotic proteins of the Bcl 2 family genes. The pro-apoptotic proteins of Bcl-2 family includes Bad, Bik and Bid and the anti-apoptotic proteins include Bcl-2[43] and Bcl-xL [44]. Formation of homodimers of Bcl-2 in mitochondrial membrane prevents the activation of caspase-9 while formation of heterodimers of Bcl-2 and Bad activates caspase-9[45]. Regulation of apoptotic pathway by Akt involves the phosphorylation of Bad on Serine 136 thereby preventing the formation of heterodimers in mitochondrial membrane[46].

N-cadherin is typically expressed by mesenchymal cells which are more motile in nature than epithelial cells. Studies have reported unusual expression of N-cadherin in epithelium derived tumors and this upregulation of expression of N-cadherin promotes cell motility and invasiveness[47][48]. This shift in the expression of cadherins from E-cadherin to N-cadherin occur during gastrulation where it affects the phenotype of participating cells and helps in the separation of different types of cells, for example, a shift in expression from E-cadherin to N-cadherin helps the segregation of neural tube from the epithelium [49][50].

#### Cadherin switching and its role in prostate cancer metastasis

Cadherin switching usually refers to shifting of E-cadherin expression to N-cadherin expression but also involves conditions where N-cadherin expression is upregulated without a significant change in expression of E-cadherin and also situations where other cadherins like R-cadherin, P-cadherin, T-cadherin and cadherin-11 etc, are co expressed with E-cadherin[51][52][53]. Cadherin switching has been reported to be an important event occurring during metastatic progression of a tumor by enhancing the invasiveness of the tumor cells[54][55][56][57]. Decrease in expression of E-cadherin and increase in N-cadherin expression has been observed in various metastatic tumors. Studies on prostate cancer cell lines have also reported upregulation of N-cadherin expression that might mediate a homotypic adhesion between prostate cancer cells and stromal fibroblasts and facilitate metastasis[58]. Invasion of prostate cancer proceed through the surrounding stroma, migration to the perineural space and finally

penetrate the capsule to escape from the primary location[59][60]. In addition to facilitating the escape from prostate gland N-cadherin expression might also aid the invasion of local blood vessels by the tumor cells. As endothelial cells also express N-cadherin in extra-junctional spaces, with an unclear role[61], a homotypic interaction between prostate cancer cells and endothelial cells promote metastasis by allowing access to the blood vascular system, possibly involving the IL-6-TGF- $\beta$ -MMP-9 pathway, as demonstrated by *ex vivo* cell culture experiments[62]. Cadherin switching, decreased expression of E-cadherin and increased expression of N-cadherin, was observed in LNCaP-19 tumor cells, as the tumor progressed towards a stage of androgen independency suggestive of a correlation between cadherin switching, invasiveness and androgen independency in prostate cancer[63]. Studies have shown that apoptosis is induced in both normal and cancer cells, when cadherin adhesion is disrupted[64]. Also, increased Akt expression has been observed in androgen-independent metastatic prostate cancer cells[65].

In most of the epithelial malignancies, a key step in metastasis of carcinomas of breast and prostate is the transcriptional repression of E-cadherin gene[66]. Although the mechanisms which regulate the abnormal expression of N-cadherin in carcinoma progression are yet unknown, it has been shown that N-cadherin expression during epithelial-mesenchymal transition is induced by TGF $\beta$ 1 through GTPase RhoA signalling[67] while at the later stages, prostate cancer cells are resistant to this growth factor[68]. A basic helix-loop-helix transcription factor Twist-1 which regulates the expression of E-cadherin and increased expression of mesenchymal genes during morphogenesis has been shown to be up-regulated in breast and prostate carcinomas[69].

A study on the role of Twist-1 in regulating N-cadherin expression has shown that increased accumulation of Twist-1 in nucleus results in  $\beta$ 1 integrin mediated cell adhesion. Twist-1 directly binds to an E-box cis-element located in the first intron of the human N-cadherin gene and initiates the transcription of N-cadherin[70]. N-cadherin also plays dual functional roles in homophilic cell-cell adhesion and regulation of apoptosis. Studies involving PC3 cell lines have shown that homophilic adhesion of N-cadherin is linked to Akt signalling and inhibition of mitochondrial apoptotic pathway. Homophilic adhesion between extracellular domains of N-cadherin provides specific signals that regulate the levels of Bcl-2 by recruitment and activation of PI3-kinase and phosphorylation of Akt which leads to phosphorylation of Bad at Ser-136 and stabilizes Bcl-2[71].

#### Role of ZEB1 in promoting EMT in prostate cancer cells

Zinc finger enhancer binding protein (ZEB1) is a zinc finger homeo-domain transcription repressor and is known to regulate developmental processes like muscle, lymphoid differentiation and skeletal patterning[72]. ZEB1 expression has also been shown to be elevated in various malignancies like breast, lung and colorectal cancer [73][74][75]. It represses the expression of E-cadherin by interacting with CANNTG sequence in the promoter region and recruiting histone deacetylase, thereby resulting in chromatin condensation and gene silencing[72][73]. *In-vitro* studies to investigate the relationship between expression of ZEB1 and prostate cancer, using metastatic prostate cancer cell lines DU-145, PC-3, ARCaP<sub>E</sub> and ARCaP<sub>M</sub> and poorly tumorigenic cell line LNCaP as well as its bone- derived sub line C4-2B, has shown that ZEB1 mRNA and protein expression is undetectable in normal prostate cells, moderately expressed in low Gleason score tumors and highly expressed in tumors with high Gleason score, suggesting its relation with aggressiveness and grade of tumor.

Expression of ZEB1 is dependent on MEK signaling as the inhibition of MEK/ERK reduces the expression of ZEB1 in ARCaP<sub>E</sub> cell lines. Inhibition of MEK/ERK also suppresses the expression of  $\beta$ -catenin (also reported to regulate cadherin-11), however, it does not have any effect on the expression of endogenous mesenchymal markers like fibronectin and vimentin. Inhibition of ZEB1 by using ZEB1-siRNA revealed decreased migration rate in otherwise aggressive ARCaP<sub>M</sub> cell lines suggesting that ZEB1 also plays a role in suppressing the expression of E-cadherin and promoting the expression of N-cadherin[76]. Specifically, it has been observed that

nicotinamide adenine dinucleotide (NAD)-dependent histone deacetylase (SIRT1) deacetylates histone H3, following recruitment to the E-cadherin proximal promoter by ZEB1. This, in turn, reduces the binding of RNA polymerase II to the transcriptional start site, ultimately suppressing E-cadherin transcription[77]. However, other experiments have demonstrated that exit from EMT involves an up-regulation of E-cadherin, despite the persistent expression of ZEB1 providing evidence for the need to use an appropriate model system for attempts to replicate E-cadherin expression in human cancers[78].

Expression of IGF is also correlated with the expression of ZEB1 in the serum of patients with high Gleason Score Prostate cancer, suggesting a role of IGF1 signaling in the over-expression of ZEB1. Highly aggressive cell lines like ARCaP<sub>M</sub> has a 2 fold increase in the expression of phosphorylated IGF-IR $\beta$ [79]. Further, the expression of ZEB1 in MEK/ERK suppressed cell lines is restored on treatment with IGF-1 (a factor that also induces the expression of Twist and is known to promote EMT)[80]. While corroborative data indicate the involvement of IGF-1, via ZEB1, in prostate cancer initiation, a large study (ProtecT trial) which evaluated the relationship between circulating insulin-like growth factors (IGFs) and prostate cancer has found no role for circulating serum IGF-1 with reference to this cancer[81]. However, more recent evidence seems to indicate that a better evaluation of the nature of the relationship may be done by measuring the levels of distinct isoforms during the progression of prostate cancer[82] with suitable and acceptable histo-pathological correlates.

#### Role of snail, slug and E-47 factors in EMT and metastasis

Zinc finger factors Snail, Slug and basic Helix-Loop-Helix factor E-47, like ZEB1, also plays an important role in EMT. These factors, when expressed, induce a similar kind of phenotype which exhibit complete EMT[83][84][85]. Gene profiling studies involving MDCK-Snail, MDCK-Slug and MDCK-E-47 cell lines have shown an up-regulation of transcription factor IGF-1, cell proliferation and signaling factor Wnt-5 and various other genes related to EMT, angiogenesis, metabolism, transport and basic cellular functions. About 36% of the EMT-related genes were coordinately regulated by all the 3 genes, while the remaining was regulated by one or more of the 3 afore-said transcription factors. Regulation of over-expressed EMT-related genes in these cell lines is in a similar fashion by either of these factors provides evidence for their important role in EMT and imparting metastatic & invasive potential to tumorous cells[86].

Apart from the insulin-like growth factors, hepatocyte growth factor[87], epidermal growth factor[88], fibroblast growth factor[89], the transforming growth factor[90] plays an important role in EMT processes [91]. miR-200 and miR-205 have been recently shown to modulate the function of ZEB1 and ZEB2 (transcriptional repressors of E-cadherin gene expression), thereby playing an important role in TGF $\beta$ -induced EMT[92].

#### Role of TGF $\beta$ and oncogenic ras in EMT and metastasis

Oncogenic Ras (normally mitogenic) and Transforming Growth Factor- $\beta$  (TGF $\beta$ ) (normally growth inhibitory) and TGF $\beta$  Receptor are known to play important roles in EMT and metastasis. TGF $\beta$ R signaling is required for EMT, invasion and metastasis in cancer cell through a Rho-dependent mechanism as mentioned earlier[93][94][95]. However, controversial to this is the known, paradoxical role of TGF $\beta$  in tumor suppression by growth inhibition and it functions as a tumor suppressor gene[96]. Downstream signaling pathways of oncogenic Ras are complex and involve many feedback loops as well as cross-talk with other pathways. It is mediated through Raf/MEK/ERK signaling and is required for TGF $\beta$ -induced EMT and metastasis. In addition, the activation of PI3K (phosphatidylinositol 3 kinase) - another TGF $\beta$ -regulated pathway, by downstream signaling of Ras oncogene, protects the cell from TGF $\beta$  induced apoptosis, thereby suggesting that tumor metastasis and EMT depends on mutual harmony between expression of TGF $\beta$  and PI3 Kinase[97][98][99].

#### Role of Wnt in EMT and metastasis

Gene profiling studies of various cancer cell lines have revealed an up-regulation of cell proliferation and signaling factor WNT-5. Wnt (wingless type) pathway plays a central role in the development of tissues during embryonic stages. It has also been shown that abnormal activation of Wnt pathway is involved in rendering metastatic potential and invasiveness in Prostate cancer cells[100][101]. Wnt pathway participates in cell invasion, proliferation, metastasis and angiogenesis by the regulation of target Wnt genes. Earlier, it was believed that activating mutations in  $\beta$ -catenin were the dominant mechanism in activation of Wnt in cancerous cells[102] but studies have shown that despite presence of these downstream activating mutations, presence of secreted Wnt antagonists like secreted Frizzled-related protein (sFRP)family, Dickkopf (Dkk) family and Wnt inhibitory factor-1[103][104][105] can suppress Wnt signaling suggestive of an autocrine Wnt signaling involved in tumor progression[106][107]. Corroborative evidence has been provided for the role of  $\beta$ -catenin (mRNA & protein levels), at the level of the cadherin-11 3'UTR, even though  $\beta$ -catenin-independent regulation of cadherin-11 has also been observed[108].

WIF-1 has been shown to inhibit the growth of various tumors and its expression was observed to be downregulated in 64% of primary prostate cancer specimens[109][110]. Studies using PC3 cancer cell lines revealed that inhibition of WIF-1 in most prostate cancer cells is due to hyper-methylation of its promoter[111]. Ectopic expression of WIF-1 in prostate cancer cell lines results in the upregulation of epithelial markers and increase in the protein levels of E-cadherin and keratin-18 as well as downregulates mesenchymal markers N-cadherin, fibronectin and vimentin, thereby resulting in the reversal of EMT.

Thus modulation of EMT markers is associated with the inhibition of Wnt signaling by WIF-1[112]. Inhibition of Wnt signaling down regulates the expression of Slug/Twist transcription factors, which are known to promote EMT. Restoration of WIF-1 in PC3 cell lines, thus resulted in the complete reversal of EMT, by inducing the expression of epithelial markers E-cadherin and keratin-18 and suppression of mesenchymal markers N-cadherin and vimentin, suppression of cell motility by down regulation of matrix metalloproteinases-2 and 9 and down regulation of transcription factors Slug/Twist[113]. Such mechanistic insights, in addition to those provided by E-cadherin conditional knock out and cadherin-11 knock-out animals, provide opportunities for development of molecules (like dietary polyphenols), that can potentially be used for the reversal of EMT[106] and cause the induction of programmed cell death or apoptosis[114].

#### ACKNOWLEDGEMENT

The authors would like to profusely thank the Management of VIT University for their infrastructural support and their constant source of encouraging and inspiring words which made this manuscript possible.

#### REFERENCES

1. Baade PD, Youlden DR, Krnjacki LJ. International epidemiology of prostate cancer: geographical distribution and secular trends. *Mol Nutr Food Res* 2009;53:171-84.
2. Xia SJ, Cui D, Jiang Q. An overview of prostate diseases and their characteristics specific to Asian men. *Asian J Androl* 2012;14(3):458-64.
3. Pu YS, Chiang HS, Lin CC, Huang CY, Huang KH, Chen J: Changing trends of prostate cancer in Asia. *Aging Male* 2004; 7: 120-32.
4. Grönberg, H. Prostate cancer epidemiology. *The Lancet* 2003; 361: 859-864.
5. Harada M, Lida M, Yadaguchi M, Shida K. Analysis of bone metastasis of prostatic adenocarcinoma in 137 autopsy cases. *Prostate Cancer and bone metastasis*. New York : Plenum Press; 1992. p.173-182.
6. Yin JJ, Pollock CB, Kelly K. Mechanisms of cancer metastasis to the bone. *Cell Res* 2005;15:57-62.

7. Long MA. Features of unusual metastases from prostate cancer. *The British Journal of Radiology* 1999; 72:933-941.
8. Wang CY, Wu GY, Shen MJ, Cui KW, Shen Y. Comparison of distribution characteristics of metastatic bone lesions between breast and prostate carcinomas. *Oncol Lett* 2013;5:391-397.
9. Bubendorf L, Schöpfer A, Wagner U, Sauter G, Moch H, Willi N, Gasser TC, Mihatsch MJ. Metastatic patterns of prostate cancer: an autopsy study of 1,589 patients. *Hum Pathol* 2000;31:578-83.
10. Batson OV. The function of vertebral veins and their role in the spread of metastasis. *Ann Surg* 1940; 112:138-149.
11. Batson, O.V. The function of vertebral veins in the metastatic processes. *Ann Intern Med* 1942; 16:138-145.
12. Fidler IJ, Poste, G. The "seed and soil" hypothesis revisited. *The Lancet Oncology* 2008;9:808.
13. Langley RR, Fidler IJ.: The seed and soil hypothesis revisited - the role of tumor-stroma interactions in metastasis to different organs. *Int J Cancer* 2011;128: 2527-35.
14. Koeneman KS, Yeung F, Chung LW. Osteomimetic properties of prostate Cancer cells: a hypothesis supporting the predilection of prostate cancer metastasis and growth in the bone environment. *Prostate* 1999;39:246-61.
15. Roato I. Interaction among cells of bone, immune system, and solid tumors leads to bone metastases. *Clin Dev Immunol* 2013;2013:1-7.
16. Fidler IJ. The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. *Nature Rev Cancer* 2003; 3:453-58.
17. Langley RR, Fidler IJ. Tumor cell-organ microenvironment interactions in the pathogenesis of cancer metastasis. *Endocr Rev* 2007;28:297-321.
18. Tarin D, Thompson EW, Newgreen DF. The fallacy of epithelial mesenchymal transition in neoplasia. *Cancer Res* 2005;65:5996-6000.
19. Creighton CJ, Gibbons DL, Kurie JM. The role of epithelial-mesenchymal transition programming in invasion and metastasis: a clinical perspective. *Cancer Manag Res* 2013; 5:187-95.
20. van der Pluijm G. Epithelial plasticity, cancer stem cells and bone. *Metastasis formation. Bone* 2011;48:37-43.
21. Wheelock MJ, Johnson KR. Cadherin mediated cellular signalling. *Curr Opin Cell Biol* 2003; 15:509-514.
22. Wheelock MJ, Johnson KR. Cadherins as modulators of cellular phenotype. *Annual Rev Cell Dev Biol* 2003;19:207-235.
23. Wheelock MJ, Shintani Y, Maeda M, Fukumoto Y, Johnson KR. Cadherin switching. *J Cell Sci* 2008; 121:727-35.
24. Alonso SR, Tracey L, Ortiz P, Pérez-Gómez B, Palacios J, Pollán M. et al. A high-throughput study in melanoma identifies epithelial-mesenchymal transition as a major determinant of metastasis. *Cancer Res* 2007; 67:3450-60.
25. Turley EA, Veiseh M, Radisky DC, Bissell MJ. Mechanisms of disease: epithelial mesenchymal transition--does cellular plasticity fuel neoplastic progression? *Nat Clin Pract Oncol* 2008; 5:280-90.
26. Gumbiner BM. Regulation of cadherin-mediated adhesion in morphogenesis. *Nat Rev Mol Cell Biol* 2005; 6: 622-634.
27. Patel SD, Chen CP, Bahna F, Honig B, Shapiro L. Cadherin mediated cell-cell adhesion; sticking together as a family. *Curr Opin Struct Biol* 2003; 13:690-698.
28. Brasch J, Harrison OJ, Honig B, Shapiro L. Thinking outside the cell: how cadherins drive adhesion. *Trends Cell Biol* 2012; 22:299-310.
29. Takeichi M. The cadherins: cell-cell adhesion molecules controlling animal morphogenesis. *Development* 1988; 102:639-55.
30. Takeichi M. Self-organization of animal tissues: cadherin-mediated processes. *Dev Cell* 2011; 21:24-26.
31. Troyanovsky S. Cadherin dimers in cell-cell adhesion. *Eur J Cell Biol* 2005; 84:225-33.
32. Troyanovsky RB, Laur O, Troyanovsky SM. Stable and unstable cadherin dimers: mechanisms of formation and roles in cell adhesion. *Mol Biol Cell* 2007;18:4343-52.
33. Tran NL, Nagle RB, Cress AE, Heimark RL. N-Cadherin expression in human prostate carcinoma cell lines. An epithelial-mesenchymal transformation mediating adhesion with Stromal cells. *Am J Pathol* 1999;155:787-98.
34. Chu K, Cheng CJ, Ye X, Lee YC, Zurita AJ, Chen DT. et al. Cadherin-11 promotes the metastasis of Prostate cancer cells to bone. *Mol Cancer Res* 2008;6:1259-67.
35. Johnson E, Theisen CS, Johnson KR, Wheelock MJ. R-cadherin influences cell motility via Rho family GTPases. *J Biol Chem* 2004;279:31041-49.
36. Battle E, Sancho E, Franci C, Dominguez D, Monfar M, Baulida J. et al. The transcription factor snail is a repressor of E-cadherin expression in epithelial tumor cells. *Nat Cell Biol* 2000; 2:84-89.
37. Comijin J, Berx G, Vermassen P, Verscheren K, van Grunsven L, Bruyneel F. et al. The two handed E-box binding zinc finger protein SIP1 down regulates E-cadherin and induces invasion. *Mol. Cell* 2001; 7: 1267-78.
38. Fan L, Wang H, Xia X, Rao Y, Ma X, Ma D. et al. Loss of E-cadherin promotes prostate cancer metastasis via upregulation of metastasis-associated gene 1 expression. *Oncol Lett* 2012;4:1225-33.
39. Arregui C, Pathre P, Lilien J, Balsamo J. The non-receptor tyrosine kinase fer mediates cross-talk between N-cadherin and  $\beta$ 1-integrins. *J Cell Biol* 2000;149:1263-74.
40. Weiner JA, Jontes JD. Proto-cadherins, not prototypical: a complex tale of their interactions, expression, and functions. *Front Mol Neurosci* 2013;6:1-10
41. Kovacs EM, Ali RG, McCormack AJ, Yap AS. E-cadherin homophilic ligation directly signals through Rac and phosphatidylinositol 3-kinase to regulate adhesive contacts. *J. Biol. Chem* 2002; 277: 6708-6718.
42. Persad S, Attwell S, Gray V, Mawji N, Deng JT, Leung D. et al. *J. Biol. Chem* 2001; 276:27462-27469.
43. Bojes HK, Suresh PK, Mills EM, Spitz DR, Sim JE, Kehrer JP. Bcl-2 and Bcl-xL in peroxide-resistant A549 and U87MG cells. *Toxicol Sci* 1998 Apr;42(2):109-16.
44. Adams JM, Cory S. The Bcl-2 Protein Family: Arbiters of Cell Survival *Science* 1998; 281:1322-1326.
45. Green DR, Reed JC. Mitochondria and apoptosis. *Science* 1998; 281:1309-12.
46. Datta SR, Dudek H, Tao X, Masters S, Fu H, Gotoh Y. et al. *Cell* 1997; 91:231-241.
47. Hazan RB, Phillips GR, Qiao RF, Norton L, Aaronson SA. Exogenous expression of N-cadherin in breast cancer cells induces cell migration, invasion and metastasis. *J. Cell. Biol* 2000;148: 779-790.
48. Gravdal K, Halvorsen OJ, Haukaas SA, Akslein LA. A switch from E-cadherin to N-Cadherin expression indicates epithelial to mesenchymal transition and is of strong and independent importance for the progress of Prostate Cancer. *Cancer Res* 2007; 13: 7003-7011.
49. Cavallaro U, Schaffhauser B, Christofori G. Cadherins and the tumor progression: is it all in a switch? *Cancer Lett* 2002; 176:123-128.
50. Christofori G. Changing neighbours, changing behaviour; cell-adhesion molecule mediated signalling during tumor progression. *EMBO J* 2003; 22:2318-2323.
51. Tomita K, van Bokhoven A, van Leenders GJ, Ruijter ET, Jansen CF, Bussemakers MJ. et al. Cadherin switching in human prostate cancer progression. *Cancer Res* 2000; 60: 3650-3654.
52. Derycke LD, Bracke ME. N-cadherin in the spotlight of cell-cell adhesion, differentiation, embryogenesis, invasion and signalling. *Int J Dev Biol* 2004; 48:463-476.
53. Tamiuchi K, Nakagawa H, Hosokawa M, Nakamura T, Eguchi H, Ohigashi H, et al. Overexpressed P-cadherin/CDH3 promotes motility of pancreatic cancer cells by interacting with p120 catenin and activating rho-family GTPases. *Cancer Res* 2005; 65:3092-3099.
54. Hult J, Suyama K, Chung S, Keron R, Agiostratidois G, Shan W. et al. N-cadherin signalling potentiates mammary tumor metastasis via enhanced extra cellular signal-related kinase activation. *Cancer Res* 2007; 67:3106-3116.
55. Knudsen KA, Sauer C, Johnson KR, Wheelock MJ. Effect of N-cadherin misexpression by the mammary epithelium in mice. *J. Cell Biochem* 2005; 95:1093-1107.
56. Riou P, Saffroy R, Chenailler C, Franc B, Gentile C, Rubinstein E, et al. Expression of T-cadherin in tumor cells influences

- invasive potential of human hepatocellular carcinoma. *FASEB J* 2006; 20:2291-2301.
57. Stefansson IM, Salvesen HB, Akslen, LA. Prognostic impacts of alterations in P-cadherin expression and related cell adhesion markers in endometrial cancer. *J Clin Oncol* 2004; 22:1242-1252.
  58. Tran NL, Nagle RB, Cress AE, Heimark RL. N-Cadherin expression in human prostate carcinoma cell lines. An epithelial-mesenchymal transformation mediating adhesion with stromal cells. *Am J Pathol* 1999;155:787-98.
  59. Villers A, MvNeal JE, Redwine EA, Feriha FS, Stamey TA. The role of perineural space invasion in local spread of prostatic adenocarcinoma. *J Urol* 1989; 142: 763-768.
  60. McNeal JE, Villers AA, Redwine EA, Feriha FS, Stamey, TA. Capsular penetration in prostate cancer: Significance for natural history and treatment. *Am J Surg Pathol* 1990; 14: 240-247.
  61. Novarro, P., Ruco, L. and Dejana, E. Differential localization of V-E and N-cadherin in human endothelial cells: V-E cadherin competes with N-cadherin for junctional localization. *J Cell Biol* 1998; 140: 1475-1484.
  62. Wang X, Lee SO, Xia S, Jiang Q, Luo J, Li L. et al. Endothelial cells enhance prostate cancer metastasis via IL-6→androgen receptor→TGF-β→MMP-9 signals. *Mol Cancer Ther* 2013;12:1026-37.
  63. Jennbacken K, Gustavsson, H., Welen, K., Vallbo, C. and Damber, J.E. Prostate cancer progression into androgen independency is associated with alterations in cell adhesion and invasivity. *The Prostate* 2006; 66:1631-1640.
  64. Nightingale J, Chaudhary KS, Abel PD, Stubbs AP, Romanska HM, Mitchell SE. et al. Ligand activation of the androgen receptor downregulates E-cadherin-mediated cell adhesion and promotes apoptosis of prostatic cancer cells. *Neoplasia* 2003;5:347-61.
  65. Graff JR, Konicek BW, McNulty AM, Wang Z, Houck K, Allen S. et al. Increased AKT activity contributes to prostate cancer progression by dramatically accelerating prostate tumor growth and diminishing p27Kip1 expression. *J Biol Chem* 2000;275:24500-05.
  66. Peinado H, Portillo F, Cano A. Transcriptional regulation of cadherins during development and carcinogenesis. *Int. J Dev Biol* 2004; 48:365-375.
  67. Guo Y, Kyprianou N. Overexpression of transforming growth factor (TGF) beta1 type II receptor restores TGF-beta1 sensitivity and signaling in human prostate cancer cells. *Cell Growth Differ* 1998; 9:185-93.
  68. Bhowmick NA, Ghiassi M, Bakin A, Aakre M, Lundquist CA, Engel ME. et al. Transforming growth factor-beta1 mediates epithelial to mesenchymal transdifferentiation through a RhoA-dependent mechanism. *Mol Biol Cell* 2001 Jan;12(1):27-36.
  69. Yang J, Mani SA, Donaher JL, Ramaswamy S, Itzykson RA, Come C, et al. Twist, a master regulator of morphogenesis, plays an essential role in tumor metastasis. *Cell*. 2004;117:927-939.
  70. Alexander NR, Tran NL, Rekapally H, Summers CE, Glackin C, Heimark RL. N-cadherin gene expression in Prostate carcinoma is modulated by integrin-dependent translocation of Twist-1. *Cancer Res* 2006; 66:3365-3369.
  71. Tran NL, Adams DG, Vaillancourt RR, Heimark RL. Signal transduction from N-cadherin increases Bcl-2. Regulation of the phosphatidylinositol 3-kinase/Akt pathway by homophilic adhesion and actin cytoskeletal organization. *J Biol Chem* 2002;277:32905-14.
  72. Creighton CJ, Gibbons DL, Kurie JM. The role of epithelial-mesenchymal transition programming in invasion and metastasis: a clinical perspective. *Cancer Manag Res* 2013;5:187-95.
  73. Eger A, Aigner K, Sonderegger S, Dampier B, Oehler S, Schreiber M. et al. DeltaF1 is a transcriptional repressor of E-cadherin and regulates epithelial plasticity in breast cancer cells. *Oncogene* 2005; 24:2375-85.
  74. Spoelstra NS, Manning NG, Higashi Y, Darling D, Singh M, Shroyer KR, et al. The transcription factor ZEB1 is aberrantly expressed in aggressive uterine cancers. *Cancer Res* 2006;66:3893-902.
  75. Spaderna S, Schmalhofer O, Hlubek F, Bex G, Eger A, Merkel S, et al. A transient, EMT-linked loss of basement membranes indicates metastasis and poor survival in colorectal cancer. *Gastroenterology* 2006;131:830-40.
  76. Grootclaes M, Frisch S. Evidence for a function of CtBP in epithelial gene regulation and anoikis. *Oncogene* 2000;19:3823-28.
  77. Chinnadurai G. CtBP, an unconventional transcriptional corepressor in development and oncogenesis. *Mol Cell* 2002;9:213-24.
  78. Graham TR, Zhou HE, Odero-Marrah VA, Osunkoya AO, Kimbro KS, Tighiouart M, et al. Insulin-like growth factor-I-dependent up-regulation of ZEB1 drives epithelial-to-mesenchymal transition in human prostate cancer cells. *Cancer Res* 2008;68(7):2479-88.
  79. Byles V, Zhu L, Lovaas JD, Chmielewski LK, Wang J, Faller DV. et al. SIRT1 induces EMT by cooperating with EMT transcription factors and enhances prostate cancer cell migration and metastasis. *Oncogene* 2012;31:4619-29.
  80. Putzke AP, Ventura AP, Bailey AM, Akture C, Opoku-Ansah J, Celiktaş M. et al. Metastatic progression of prostate cancer and e-cadherin regulation by Zeb1 and SRC family kinases. *Am J Pathol*. 2011; 179:400-10.
  81. Kwok WK, Ling MT, Lee TW, Lau TC, Zhou C, Zhang X. et al. Up-regulation of TWIST in prostate cancer and its implication as a therapeutic target. *Cancer Res* 2005; 65:5153-62.
  82. Rowlands MA, Holly JM, Gunnell D, Gilbert R, Donovan J, Lane JA. et al. The relation between adiposity throughout the life course and variation in IGFs and IGFs: evidence from the ProtecT Prostate testing for cancer and Treatment) study. *Cancer Causes Control* 2010;21:1829-42
  83. Philippou A, Armakolas A, Koutsilieris M. Evidence for the Possible Biological Significance of the IGF-1 Gene Alternative Splicing in Prostate Cancer. *Front Endocrinol (Lausanne)* 2013;4:1-11.
  84. Cano A, Pérez-Moreno MA, Rodrigo I, Locascio A, Blanco MJ, del Barrio MG et al. The transcription factor snail controls epithelial-mesenchymal transitions by repressing E-cadherin expression. *Nat Cell Biol* 2000;2:76-83.
  85. Bolos V, Peinado H, Perez-Moreno MA, Fraga MF, Esteller M, Cano A. The transcription factor Slug represses E-cadherin expression and induces epithelial to mesenchymal transitions: a comparison with Snail and E47 repressors. *J Cell Sci* 2003;116:499-511.
  86. Perez-Moreno MA, Locascio A, Rodrigo I, Dhondt G, Portillo F, Nieto MA. et al. A new role for E12/E47 in the repression of E-cadherin expression and epithelial-mesenchymal transitions. *J Biol Chem* 2001;276:27424-31.
  87. Moreno-Bueno G, Cubillo E, Sarrió D, Peinado H, Rodríguez-Pinilla SM, Villa S. et al. Genetic profiling of epithelial cells expressing E-cadherin repressors reveals a distinct role for Snail, Slug, and E47 factors in epithelial-mesenchymal transition. *Cancer Res* 2006; 66:9543-9556.
  88. Oft M, Peli J, Rudaz C, Schwarz H, Beug H, Reichmann E. TGFβ1 and Ha-Ras collaborate in modulating the phenotypic plasticity and invasiveness of epithelial tumor cells. *Genes Dev* 1996; 10:2462-2477.
  89. Savagner P, Yamada KM, Thiery, JP. The zinc finger protein slug causes desmosome dissociation, an initial and necessary step for growth factor-induced epithelial-mesenchymal transition. *J Cell Biol* 1997; 137:1403-1419.
  90. Lo HW, Hsu SC, Xia W, Cao X, Shih, JY, Wei, Y. Epidermal growth factor receptor cooperates with signal transducer and activator of transcription 3 to induce epithelial-mesenchymal transition in cancer cells via up-regulation of TWIST gene expression. *Cancer Res* 2007;67:9066-9076.
  91. Lee JM, Dedhar S, Kalluri R, Thompson EW. The epithelial-mesenchymal transition: new insights in signaling, development, and disease. *J Cell Biol* 2006; 172: 973-981.
  92. Zavadil J, Bottinger, EP. TGF-beta and epithelial-to-mesenchymal transitions. *Oncogene* 2005, 24, 5764-5774.
  93. Gregory, PA, Bert AG, Paterson EL, Barry SC, Tsykin A, Farshid, G. The miR-200 family and miR-205 regulate epithelial to

- mesenchymal transition by targeting ZEB1 and SIP1. *Nat Cell Biol* 2008;10: 593-601.
94. Oft, M, Heider, KH, Beug H. TGF $\beta$  signaling is essential for carcinoma cell invasiveness and metastasis. *Curr Biol* 1998;8:1243-1252.
  95. Parsons R, Myeroff, LL, Liu B, Willson, JK, Markowitz, SD, Kinzler, KW, Vogelstein, B. Microsatellite instability and mutations of the transforming growth factor beta type II receptor gene in colorectal cancer. *Cancer Res* 1995; 55:5548-5550.
  96. Janda E, Lehmann K, Killisch I, Jechlinger M, Herzig M, Downward J, et al. Ras and TGF[ $\beta$ ] cooperatively regulate epithelial cell plasticity and metastasis: dissection of Ras signaling pathways. *J Cell Biol* 2002; 156:299-313.
  97. Castellano E, Downward J. RAS Interaction with PI3K: More Than Just Another Effector Pathway. *Genes Cancer* 2011;2:261-74.
  98. Emami KH, Corey E. When prostate cancer meets bone: control by Wnts. *Cancer Lett* 2007; 253:170-179.
  99. Paul S, Dey A. Wnt signaling and cancer development: therapeutic implication. *Neoplasma* 2008;55:165-76.
  100. Hajra KM, Fearon ER. Cadherin and catenin alterations in human cancer. *Genes Chromosomes Cancer* 2002; 34:255-268.
  101. Hoang B, Moos M Jr, Vukicevic S, Luyten FP. Primary structure and tissue distribution of FRZB, a novel protein related to *Drosophila* frizzled, suggest a role in skeletal morphogenesis. *J Biol Chem* 1996; 271:26131-37.
  102. Leyns L, Bouwmeester T, Kim SH, Piccolo S, De Robertis EM. Frzb-1 is a secreted antagonist of Wnt signaling expressed in the Spemann organizer. *Cell* 1997; 88:747-56.
  103. Glinka A, Wu W, Delius H, Monaghan AP, Blumenstock C, Niehrs C. Dickkopf-1 is a member of a new family of secreted proteins and functions in head induction. *Nature* 1998; 391:357-62.
  104. Farina AK, Bong YS, Feltes CM, Byers SW. Post-transcriptional regulation of cadherin-11 expression by GSK-3 and  $\beta$ -catenin in prostate and breast cancer cells. *PLoS One* 2009;4:1-9.
  105. Bafico A, Liu G, Goldin L, Harris V, Aaronson SA. An autocrine mechanism for constitutive Wnt pathway activation in human cancer cells. *Cancer Cell* 2004;6:497-506.
  106. Wissmann C, Wild PJ, Kaiser S, Roepcke S, Stoehr R, Woenckhaus M. et al. WIF1, a component of the Wnt pathway, is downregulated in prostate, breast, lung, and bladder cancer. *J Pathol* 2003; 201:204-212.
  107. Suzuki H, Watkins DN, Jair KW, Schuebel KE, Markowitz SD, Chen WD. et al. Epigenetic inactivation of SFRP genes allows constitutive WNT signaling in colorectal cancer. *Nat Genet* 2004; 36:417-22.
  108. Ohgashi T, Mizuno R, Nakashima J, Marumo K, Murai M. Inhibition of Wnt signaling downregulates Akt activity and induces chemosensitivity in PTEN-mutated prostate cancer cells. *Prostate* 2005;62:61-68.
  109. Yee DS, Tang Y, Li X, Liu Z, Guo Y, Ghaffar S, et al. The Wnt inhibitory factor 1 restoration in prostate cancer cells was associated with reduced tumor growth, decreased capacity of cell migration and invasion and a reversal of epithelial to mesenchymal transition. *Mol. Cancer* 2010; 9:162.
  110. Link A, Balaguer F, Goel A. Cancer chemoprevention by dietary polyphenols: promising role for epigenetics. *Biochem Pharmacol* 2010; 80:1771-92.
  111. Kumar S, Suresh PK, Vijayababu MR, Arunkumar A, Arunakaran J. Anticancer effects of ethanolic neem leaf extract on prostate cancer cell line (PC-3). *J Ethnopharmacol* 2006;105:246-50.